

Research



Cite this article: Allan BJM, Domenici P, Watson SA, Munday PL, McCormick MI. 2017 Warming has a greater effect than elevated CO₂ on predator–prey interactions in coral reef fish. *Proc. R. Soc. B* **284**: 20170784. <http://dx.doi.org/10.1098/rspb.2017.0784>

Received: 11 April 2017

Accepted: 30 May 2017

Subject Category:

Behaviour

Subject Areas:

behaviour, ecology, cognition

Keywords:

climate change, predator–prey interactions, coral reef fish, interacting stressors

Author for correspondence:

Bridie J. M. Allan

e-mail: bridie.allan@my.jcu.edu.au

Warming has a greater effect than elevated CO₂ on predator–prey interactions in coral reef fish

Bridie J. M. Allan^{1,2}, Paolo Domenici³, Sue Ann Watson¹, Philip L. Munday¹ and Mark I. McCormick^{1,2}

¹ARC Centre of Excellence for Coral Reef Studies, and ²Department of Marine Biology and Aquaculture, College of Science and Engineering, James Cook University, Townsville, Queensland 4811, Australia

³CNR-IAMC, Istituto per l'Ambiente Marino Costiero, Località Sa Mardini, 09170 Torregrande (Oristano), Italy

BJMA, 0000-0002-5991-9711; PD, 0000-0003-3182-2579; SAW, 0000-0002-9818-7429

Ocean acidification and warming, driven by anthropogenic CO₂ emissions, are considered to be among the greatest threats facing marine organisms. While each stressor in isolation has been studied extensively, there has been less focus on their combined effects, which could impact key ecological processes. We tested the independent and combined effects of short-term exposure to elevated CO₂ and temperature on the predator–prey interactions of a common pair of coral reef fishes (*Pomacentrus wardi* and its predator, *Pseudochromis fuscus*). We found that predator success increased following independent exposure to high temperature and elevated CO₂. Overall, high temperature had an overwhelming effect on the escape behaviour of the prey compared with the combined exposure to elevated CO₂ and high temperature or the independent effect of elevated CO₂. Exposure to high temperatures led to an increase in attack and predation rates. By contrast, we observed little influence of elevated CO₂ on the behaviour of the predator, suggesting that the attack behaviour of *P. fuscus* was robust to this environmental change. This is the first study to address how the kinematics and swimming performance at the basis of predator–prey interactions may change in response to concurrent exposure to elevated CO₂ and high temperatures and represents an important step to forecasting the responses of interacting species to climate change.

1. Introduction

Atmospheric CO₂ has risen from approximately 280 ppm pre-industrially to 400 ppm in 2016, the highest level in at least 800 000 years [1]. CO₂ levels in the ocean are rising at the same rate as in the atmosphere, leading to a decline in seawater pH [2]. If the current rate of anthropogenic CO₂ emissions is maintained, climate models project that the partial pressure of CO₂ (*p*CO₂) in the surface ocean will exceed 900 μatm by the end of this century and ocean pH will decline by 0.14–0.43 units compared with current-day values [1,3]. These changes in ocean chemistry are set against a background of warming, with average ocean temperatures projected to increase by up to 3°C [3]. This climate forcing can lead to positive feedback loops whereby increasing temperature can amplify the release of CO₂ from terrestrial and marine sinks, leading to further increases in temperature [1]. Consequently, marine organisms face complex environmental changes throughout this century. While there has been extensive research into the effects of ocean acidification and rising temperature on marine organisms, the majority have only tested the effects of these drivers in isolation, with fewer studies exploring their combined effects, although the importance of testing for combined effects of multiple drivers is increasingly recognized [4–7]. Understanding how multiple drivers affect key physiological or behavioural traits in marine organisms is critical, as extrapolations based on single driver responses could lead to incorrect predictions about future impacts [5,7].

In marine fish, some of the most marked effects of elevated CO₂ and warming reported to date are altered behaviours and physiology. Behavioural effects of elevated CO₂ include reduced anti-predator responses, altered olfactory and auditory preferences, loss of learning, changes in activity levels, altered behavioural lateralization and visual risk assessment [6,8,9]. These behavioural changes translate into consequences for survival in natural habitats [10,11]. Similarly, higher temperature strongly influences the physiological performance of fishes leading to reductions in aerobic scope (the difference between maximum and resting metabolic rate) in some species [12,13], reduced cardiac output [14], decreased muscle development [15] and changes to the contractile properties of the swimming muscles [16] and sensory performance [17]. Similar to exposure to elevated CO₂, higher temperatures also lead to changes in behaviours such as greater activity rates [18] and sensory responsiveness [19] and increased interactions between predators and their prey [20–22].

Emerging evidence suggests that the behaviour and physiology of fishes are sensitive to changes in simultaneous exposure to CO₂ and temperature. For example, there can be effects on predation rates and predator selectivity [23], directional changes in lateralization [24], foraging behaviour [25], activity rates [26], offspring quality [27] and aerobic scope [28], all of which can influence persistence of species into the future. However, the direction of reactions to multiple stressors is not always clear. For example, if multiple stressors affect similar pathways, an additive effect may be observed [29]. By contrast, antagonistic or synergistic effects may occur if the stressors affect different pathways, depending on the direction of the response to each stressor [30].

A critical stage in the life of reef fishes occurs at the end of the larval phase, when they settle to the benthic environment. Mortality rates during the first few days of settlement are especially high. It is at this stage that fishes undergo a severe predation-induced bottleneck [31]. Success at this life stage is dependent upon the size, growth rate and fast-start performance of new recruits [32–34]. Fast starts are short, high-energy swimming bursts that are driven by the rapid contraction of the white (anaerobic) muscle fibres [35,36]. The fast kinematics of escape responses are usually controlled by the large Mauthner neurons, which are triggered as a reaction to the fast approach of a predator, although other reticulospinal cells may also be involved [37]. Successful fast starts consist of finely tuned responsiveness and locomotor performance [38]. However, fast starts are sensitive to differences in ambient temperatures and CO₂ due to the interplay between temperature and muscle contraction rates [16], and elevated CO₂ exposure and sensory performance [17]. Fast-start performance can also differ between predators and their prey due to differences in their temperature tolerance and sensitivity to elevated CO₂ [20–22].

To date, only one study has investigated the combined effects of multiple climate change drivers on predator–prey dynamics in coral reef fishes [23]. Therefore, the aim of this study was to employ a multi-stressor approach to assess the independent and combined effects of both elevated CO₂ and high temperatures on the attack and escape performance of a common pair of coral reef fishes: the piscivorous dusky dottyback (*Pseudochromis fuscus*), and its prey, the juvenile of a common damselfish (*Pomacentrus wardi*). Previous work has shown that both the predator (*Ps. fuscus*) and prey

(*Po. wardi*) are sensitive to increases in ambient CO₂ and temperature [10,20,39]. *Pomacentrus wardi* also displayed behavioural impairment following simultaneous exposure to both stressors [24]. By exposing predators and prey to elevated CO₂ and high temperatures, both independently and simultaneously, we were able to determine the relative effects of each stressor on the attack and escape performance that forms the basis of predator–prey interactions. To date, this is the first study examining how the swimming kinematics of an attacking predator and the reactivity and swimming performance of its escaping prey may change following concurrent exposure to elevated CO₂ and temperature.

2. Material and methods

(a) Study species

Pomacentrus wardi (Pomacentridae) is a small planktivorous fish commonly found on Indo-Pacific coral reefs and was chosen as our prey organism. Newly metamorphosed *Po. wardi* larvae (range 11.2–15.5 mm, average 13.2 ± 1.2 mm standard length (SL) \pm standard deviation (s.d.)) were collected using light traps moored 100 m from the fringing reef off Lizard Island in the northern Great Barrier Reef, Australia. On the morning of capture, larvae were transferred into 30 l aquaria at either control (approx. 405 μ atm) or elevated CO₂ (approx. 930 μ atm) and cross-factored with control (approx. 27°C) or high temperature (approx. 30°C). Control conditions were ambient for the study site at the time of the study (October 2012). Larvae were split into four different treatment groups in a 2 temperature \times 2 CO₂ design: control temperature–control CO₂ ($n = 17$); high temperature–control CO₂ ($n = 19$); elevated CO₂–control temperature ($n = 18$); elevated CO₂–high temperature ($n = 18$). For fish in the high (+3°C) temperature treatments, the temperature was raised by 1°C every 8 h until the final temperature of approximately 30°C was reached [20]. Following this time period, the fish were placed into three tanks per treatment where they remained for 7 days. Fish were fed four times daily ad libitum with newly hatched *Artemia* sp. but were starved for the 12 h prior to commencement of experimental trials to standardize for satiation.

The dottyback, *Ps. fuscus* (Pseudochromidae), was used as the predator. *Pseudochromis fuscus* is widely distributed throughout the Indo-Pacific and has been observed to prey upon newly settled coral reef fishes [40], including *Po. wardi*. Adult *Ps. fuscus* (range 63–99 mm, mean 78.2 ± 9.4 mm) were collected on SCUBA using a solution of clove oil diluted with ethanol and seawater from around the shallow fringing reef off Lizard Island. Immediately after collection, fish were placed within three 30 l aquaria per treatment. To avoid aggressive interactions, fish were housed individually in mesh breeding baskets (16.5 cm L \times 12.5 cm W \times 12.5 cm H) containing small lengths of PVC pipe to be used as shelter. Predators were maintained in treatment tanks for 7 days (following the same treatment exposure as *Po. wardi*) and were fed two juvenile reef fish morning and night and then not fed for the last 24 h prior to the interaction trial to standardize for satiation. A 12 L : 12 D regime was used. We used 72 *Ps. fuscus* in total: $n = 17$ for control temperature–control CO₂, $n = 19$ for high temperature–control CO₂, $n = 18$ for elevated CO₂–control temperature and $n = 18$ for high temperature–elevated CO₂.

(b) CO₂ and temperature treatments

Tanks were heated with 300 W bar heaters and insulated to ensure stability of the experimental temperatures (27 and 30°C). CO₂ treatments were maintained by CO₂ dosing to a set

pH. Seawater was pumped from the ocean into 60 l sumps where it was infused with ambient air (control) or CO₂ to achieve a pH_{NBS} of 7.87. The pH value was selected to achieve the approximate CO₂ conditions required, based on preliminary observations of total alkalinity, salinity and temperature of seawater at Lizard Island. A pH-controller (Aqua Medic, Germany) was attached to the CO₂ treatment sump to maintain pH at the desired level. A solenoid injected a slow stream of CO₂ into a powerhead at the bottom of the sump whenever the pH of the seawater rose above the set point. Equilibrated seawater from each sump was supplied at a rate of approximately 720 ml min⁻¹ to each of the aquaria. Temperature and pH_{NBS} of each aquarium were measured daily with a temperature probe (C22, Comark, Norwich, UK) and a pH meter (HQ40d, Hach, CO, USA) calibrated with fresh pH_{NBS} buffers. Seawater CO₂ in the treatment tanks was confirmed with a portable CO₂-equilibrator and non-dispersive infrared (NDIR) sensor (GMP343, Vaisala, Helsinki, Finland) [41,42]. The CO₂-equilibrator consisted of a coil of CO₂-permeable silicone tubing that was submerged in water and connected in a closed loop of CO₂ impermeable tubing to the Vaisala GMP343 sensor. Air in the closed loop was circulated between the CO₂-equilibrator and the Vaisala GMP343 by a small pump. The pCO₂ of the air in the closed loop was equilibrated with the pCO₂ in the treatment water and measured by NDIR in the Vaisala GMP343 sensor. This cross-validation confirmed that pCO₂ calculated from water chemistry (pH_{NBS} and total alkalinity) was within 5.1–56.0 μatm of pCO₂ measured by NDIR ($N = 9$). Total alkalinity of seawater was estimated by Gran titration (888 Titrando, Metrohm, Switzerland) from water samples taken twice weekly from control and treatment tanks. Alkalinity standardizations achieved accuracy within 1% of certified reference material from Dr A. Dickson (Scripps Institution of Oceanography). Seawater pCO₂ was calculated from seawater parameters in the program CO₂SYS [43] using the constants of Mehrbach *et al.* [44], refit by Dickson & Millero [45]. Seawater parameters are shown in table 1.

(c) Laboratory assays

Predator–prey interactions were evaluated using a standard protocol established by Allan *et al.* [32]. Predators and prey that had experienced the same treatment were paired together, i.e. control temperature–control CO₂-exposed predators and prey were paired together. Briefly, one predatory *Ps. fuscus* was placed into the experimental arena and one *Po. wardi* was released into an opaque PVC tube (11 cm diameter, 15 cm high) placed upright in the middle of the experimental arena (38 cm × 58 cm × 10 cm water height). Both fish were allowed to acclimate for 30 min. The PVC tube was then carefully raised and removed from the tank using a wire connected to the top of the tube; this allowed the predator and the prey to start the interaction. Trials were only used when the predator was at the opposite end of the tank to the prey at the start of the interaction to standardize for predator position. The ensuing interaction was filmed at high speed (420 fps) for 10 min or until the prey had been consumed. Maximum predator attack speed and maximum prey escape speed were measured based on the centre of mass (COM) of the fish when stretched straight based on Webb [46]. COM was assumed to be at 35% of the body length from the tip of the snout, as is the case for generalist fish [47]. Predator attacks were measured only when a predator showed a fast-directed burst towards the prey (at a speed greater than 3 body lengths s⁻¹). All variables with the exception of number of prey caught were measured using only the first attack that occurred within a trial. This was done to control for any stress either the predator or prey may have experienced due to prolonged attacks. Both predators and prey were used only once regardless of the trial outcome. Prey suffering was minimal

because prey were consumed immediately after a successful strike. Trials were conducted over 5 days in a temperature-controlled room at the same water temperature as the relevant temperature treatment. Because of the difficulties in producing enough CO₂-enriched seawater to be used in the experimental arena, control CO₂ seawater was used. Previous work has shown that the influence of CO₂ on behaviour lasts for at least 24 h after return to normal CO₂ conditions [10] and that there is no effect of using either control or elevated CO₂ water in the test arena when testing the escape performance of prey that have been exposed to control or elevated CO₂ for a similar duration to that used here [48]. Trials were randomized to account for any potential temporal or day effect.

The following variables were measured:

Predator

- (1) Capture success: percentage of trials in which the predator ingested the prey within the 10 min filming period, out of the total number of trials for each treatment.
- (2) Attack rate: number of attacks occurring within the 10 min filming period or until the prey was consumed.
- (3) Predation rate: number of captures divided by the number of attacks within the 10 min filming period.
- (4) Predator attack distance (m): the straight-line distance between the predator COM at the time the attack commenced and at the end of the attack (end is defined as when the predator came to a halt).
- (5) Predator attack speed (m s⁻¹): was measured as the average speed occurring over the attack.

Prey

- (6) Prey reaction distance (RD, measured in metres): the distance between the prey COM and the tip of the predator's snout at the onset of the escape response to a predator attack. A large RD indicates high reactivity to a predator attack.
- (7) Apparent looming threshold (ALT): the apparent looming threshold for prey escape responses to a predatory strike. This is a measure of the reactivity of the prey to the perceived threat of predation. The higher the perceived threat, the lower the ALT (in radians per second) measured at the onset of the escape response and measured as the rate of change of the angle (α) subtended by the predator's frontal profile as seen by the prey. Previous work has shown that fish tend to react to an approaching stimulus (a predator) when a given threshold of $d\alpha/dt$ (i.e. ALT) is reached. The ALT is calculated as $(4US)/(4D^2 + S^2)$, based on Dill [49] and Domenici *et al.* [50], where U is the predator speed, calculated as the speed of the predator in the frame prior to the prey's response; S is calculated based on the morphological characteristics of the predator, i.e. $S = (\text{maximal depth} + \text{maximal width})/2$, whereby both maximal depth and maximal width are at 0.25 lengths of the predator (personal observation); and D is the distance, measured at the onset of the escape response, between the prey COM and the point on the predator where its maximal width is located. Hence, $D = \text{RD} + 0.25$ lengths of the predator. For any given speed, a large RD will result in a low ALT; hence, a low ALT indicates high reactivity to a predator attack.
- (8) Directionality: this was measured by dividing escape responses into 'away' and 'towards' responses based on the first detectable movement of the head relative to the approaching predator [50].
- (9) Prey escape distance (m): the straight-line distance between the prey COM at the onset of the escape response and at the end of the escape response (i.e. when the prey came to a halt).
- (10) Mean prey escape speed (m s⁻¹): this was measured as the distance covered within a fixed time (24 ms). This fixed

Table 1. Mean (\pm s.e.) seawater parameters in the experimental system. Temperature, pH, salinity and total alkalinity (TA) were measured directly. $p\text{CO}_2$ was estimated from these parameters using CO2SYS.

CO ₂ treatment	temp treatment	temp (°C)	salinity	pH _{NBS}	total alkalinity ($\mu\text{mol kg}^{-1}$ SW)	$p\text{CO}_2$ (μatm)
control	control	26.7 (\pm 0.1)	35.2	8.18 (\pm 0.01)	2274 (\pm 6)	396 (\pm 8)
control	high	29.6 (\pm 0.1)	35.2	8.17 (\pm 0.01)	2274 (\pm 6)	415 (\pm 9)
elevated	control	26.8 (\pm 0.1)	35.2	7.87 (\pm 0.01)	2257 (\pm 6)	921 (\pm 19)
elevated	high	29.8 (\pm 0.1)	35.2	7.87 (\pm 0.01)	2257 (\pm 6)	935 (\pm 19)

duration was based on the average duration of the first two flips of the tail (22.8 ms; the first two axial bends, i.e. stages 1 and 2) defined based on Domenici & Blake [35], which is the period considered crucial for avoiding ambush predator attacks and is typical for fish within the size range used.

(d) Statistical analyses

To test the null hypothesis that capture success is independent of predator and prey temperature and elevated CO₂ exposure, capture success was compared using 4×4 contingency table analysis. Predation rate data did not meet the assumption of homogeneity of variance, so we rank transformed the data and performed a non-parametric two-way ANOVA to explore differences in predation rates between the four treatments. The effects of temperature and CO₂ exposure on performance kinematics were tested separately for prey and predators using two-factor MANOVAs with temperature and CO₂ as the fixed factors and the multiple dependent variables being: (predator) attack rate, attack distance, attack speed and (prey) RD, ALT, escape distance and escape speed. The MANOVA assumption of homogeneity of variance–covariance matrices was tested using a Box's *M* test of equality of covariance and was non-significant for the three experimental treatments (temperature, $p = 0.002$; temperature/CO₂, $p = 0.02$ and CO₂, $p = 0.3$). Univariate two-factor ANOVAs with Tukey's HSD post hoc tests were then performed to determine the nature of any differences found by the MANOVAs. Assumptions of normality and homogeneity of variance were examined using residual analysis and Levene's test. Attack rate, RD, escape speed and ALT data did not meet the assumption of normality, so we performed square root (for attack rate, RD and escape speed) and log₁₀ (for ALT) transformations, respectively, to normalize the data. Two-tailed binomial tests were performed to determine whether CO₂ or temperature exposure had an effect on the directionality of the escape response, comparing the proportion of individuals that moved away from or towards the stimulus and testing the null hypothesis of a 0.5 probability of turning either way.

3. Results

The overall capture success was significantly associated with the treatments experienced by the predator and prey ($\chi^2 = 7.82$, d.f. = 3, $p = 0.04$) (figure 1a). Capture success was greatest in the high temperature–control CO₂ treatment, with 47.3% of prey caught. Under elevated CO₂ conditions, high temperature exposure had a much-reduced effect on the probability of capture, with only 33.3% of prey captured under this treatment regime. Capture success was moderate (22%) in the elevated CO₂–control temperature treatment group and the lowest capture success occurred under current-day control temperature and control CO₂ conditions (5.8%).

The performance characteristics of the predator were unaffected by the combined effect of high temperature and elevated CO₂ (MANOVA, Pillai's trace_{3,66} = 1.3070, $p = 0.2$). There was a significant trend for predators exposed to high temperature and control CO₂ to have a higher attack rate, with more than double the attacks compared with the control temperature–control CO₂ treatment (table 2; $F_{1,67} = 4.38$, $p = 0.04$) (figure 1b). There was no effect of control temperature–elevated CO₂ and no interaction between elevated CO₂ and high temperature exposure on attack rates ($p > 0.05$). Predation rate increased following exposure to the high temperature–control CO₂ treatment, with this group displaying a marked increase in predation rate compared with the control temperature–control CO₂ treatment (table 2; $F_{1,67} = 5.99$, $p = 0.01$) (figure 1c). There was no effect of elevated CO₂ alone or an interaction effect ($p > 0.05$) on predation rate. Predator attack distance and attack speed were unaffected by the elevated CO₂ or high temperature treatments ($p > 0.05$).

The performance characteristics of prey were affected by the combined effect of high temperature and elevated CO₂ (MANOVA, Pillai's trace_{8,61} = 2.233, $p = 0.03$), and there was a significant main effect of high temperature (MANOVA, Pillai's trace_{8,61} = 3.271, $p = 0.003$). There was no multi-variate effect of elevated CO₂ independent of temperature ($p > 0.05$). Univariate two-factor ANOVAs detected significant differences between treatments in four of the tested performance attributes: two measures of reactivity (RD, whereas a large RD indicates high reactivity to a predator attack) and ALT (a high ALT value is indicative of reduced prey responsiveness) and two measures of kinematic performance (prey escape distance and prey escape speed).

There was a strong effect of high temperature on the RD of the prey (table 2; $F_{1,67} = 15.7$, $p = 0.0001$) (figure 1d). Specifically, prey exposed to high temperatures and control CO₂ allowed the predator to get twice as close to them before undertaking an escape response (0.08 m compared with 0.04 m). There was little effect of elevated CO₂–control temperature exposure on the RD of the prey, with values nearly identical to the control temperature–control CO₂ treatment (0.08 m compared with 0.075 m). When prey were exposed to the combined effect of elevated CO₂ and high-temperature treatment, RD decreased to 0.034 m; however, this was not significantly different from the other treatments.

ALT increased from 1.39 rad s⁻¹ in the control temperature–control CO₂ treatment to 14.52 rad s⁻¹ in the high temperature–control CO₂ treatment (table 2; $F_{1,67} = 16.4$, $p = 0.0001$) (figure 1e). There was a non-significant increase in ALT in response to elevated CO₂–control temperature (3.27 rad s⁻¹) and when prey were exposed to the

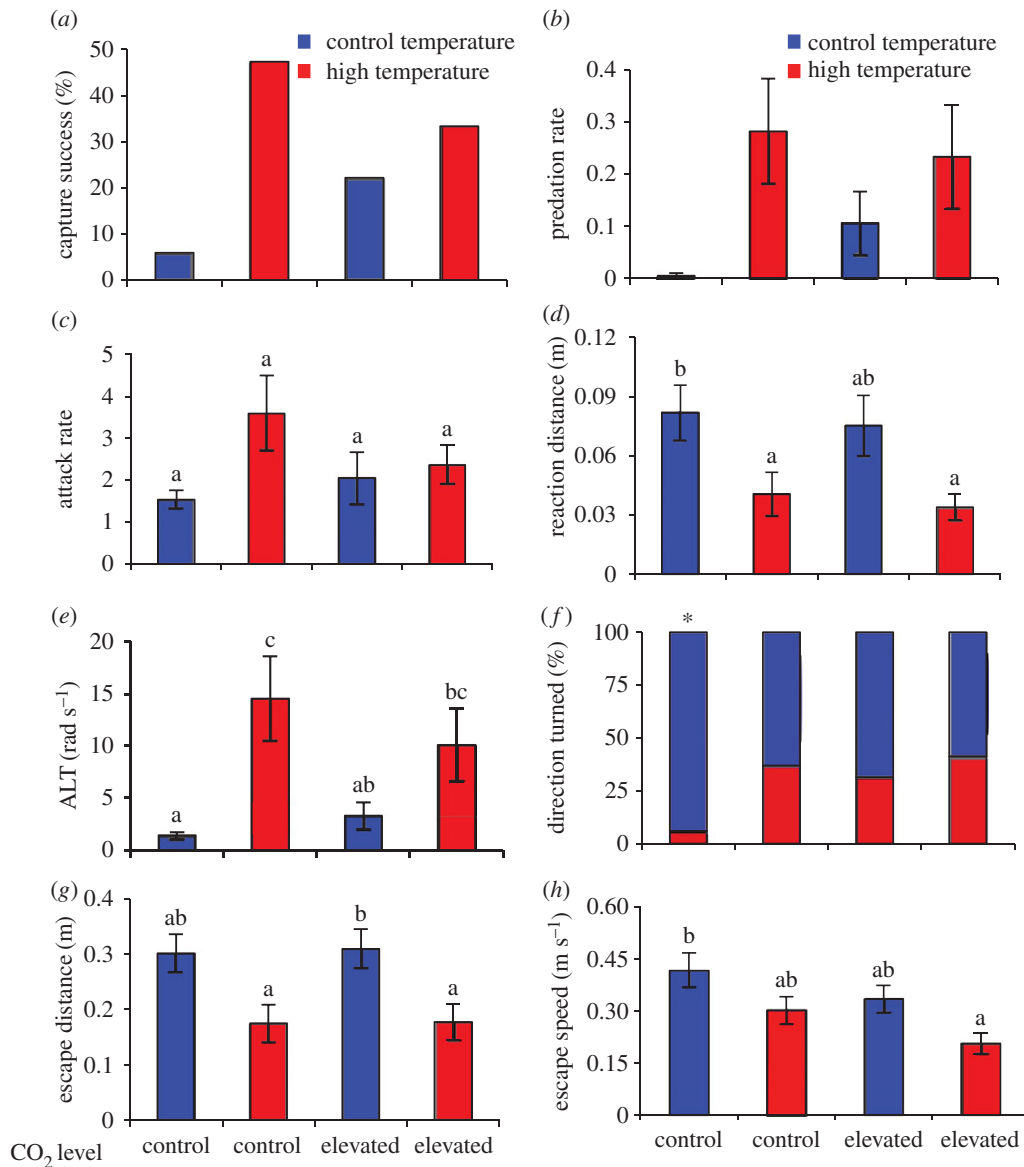


Figure 1. Comparison of the effects of elevated CO₂ (approx. 405 and 930 μatm) and temperature (control—26.7°C and elevated—29.6°C) on interactions between a predator (*Ps. fuscus*) and prey (*Po. wardi*), on eight performance characteristics: capture success (a); attack rate (within the 10 min filming period) (b); predation rate (within the 10 min filming period) (c); prey RD (d); prey ALT (e); directionality (f); prey escape distance (g); and prey escape speed (h). $n = 17$ for control temperature–control CO₂, $n = 19$ for high temperature–control CO₂, $n = 18$ for elevated CO₂–control temperature and $n = 18$ for high temperature–elevated CO₂. Error bars are s.e.m. Letters above bars represent Tukey's HSD groupings of means. Asterisk above the panel indicates that the proportion of responses directed away from (blue portion) and towards (red portion) the stimulus differed significantly from random.

combined effect of elevated CO₂–high-temperature treatment, ALT increased to 10.06 rad s^{-1} , suggestive of a strong trend. However, this was not statistically significant.

Directionality did not differ from treatments ($\chi^2 = 6.25$, d.f. = 3, $p = 0.1$) (figure 1f). However, when treatments were individually compared with the control temperature–control CO₂ treatment, there were significant differences (elevated CO₂–high temperature treatment: $\chi^2 = 9.21$, d.f. = 2, $p = 0.01$; high temperature–control CO₂, $\chi^2 = 7.37$, d.f. = 2, $p = 0.02$ and the elevated CO₂–control temperature, $\chi^2 = 5.99$, d.f. = 2, $p = 0.03$). Specifically, in the control temperature–control CO₂ treatment, most of the prey (94%) turned away from the predator. By contrast, only 58% of prey turned away from the predator following combined exposure to elevated CO₂ and high temperature. The direction turned by the prey in the high temperature–control CO₂ treatment and the elevated CO₂–control temperature treatment was intermediate to the control temperature–control CO₂ treatment and combined

effect of elevated CO₂ and high temperature treatment group. The proportion of responses directed away from and towards the predator differed significantly from random (50:50) for the control temperature–control CO₂ treatment ($n_{\text{norm}} = 17$, $p = 0.001$), but not for the other three treatments (high temperature–control CO₂: $n_{\text{norm}} = 19$, $p = 0.09$; elevated CO₂–control temperature; $n_{\text{norm}} = 16$, $p = 0.06$ or the combined effect of elevated CO₂ and high-temperature treatment: $n_{\text{norm}} = 17$, $p = 0.1$).

There was a strong effect of high temperature on the total prey escape distance (table 2; $F_{1,67} = 14.8$, $p = 0.0002$) (figure 1g), with prey exposed to high temperature–control CO₂ having 50% lower escape distances compared with prey from control temperature–control CO₂ (0.30 versus 0.17 m). By contrast, elevated CO₂ exposure–control temperature had no significant effect on prey escape distances (0.31–0.30 m; $p > 0.05$) and there was no significant interaction between the two treatments ($p > 0.05$).

Table 2. Effects of water temperature (approx. 27, 30°C) and CO₂ (approx. 405, 930 µatm) on the behavioural interaction between a predator (*Ps. fuscus*) and its prey (*Po. wardi*) on six performance variables: attack rate, predation rate, prey RD, ALT, prey escape distance and prey escape speed ($n = \text{ambient}; n = 17$; high temperature; $n = 19$; elevated CO₂; $n = 18$; high temperature and elevated CO₂; $n = 18$).

behaviour	source of variation	d.f.	MS	F	p-value
attack rate	temp	1	1.9700	4.3823	0.0401
	CO ₂	1	0.2235	0.4971	0.4832
	temp × CO ₂	1	0.5780	1.2858	0.2608
	error	67	0.4495		
predation rate	temp	1	18.282	5.992	0.0169
	CO ₂	1	0.7952	0.2606	0.6113
	temp × CO ₂	1	5.7200	1.8747	0.1754
	error	67	3.0511		
reaction distance	temp	1	0.1436	14.785	0.0002
	CO ₂	1	0.0026	0.2741	0.6023
	temp × CO ₂	1	0.0003	0.0338	0.8547
	error	67	0.0009		
ALT	temp	1	8.7191	16.402	0.0001
	CO ₂	1	0.0193	0.0363	0.8494
	temp × CO ₂	1	0.4005	0.7535	0.3884
	error	67	0.5315		
escape distance	temp	1	0.3145	14.829	0.0002
	CO ₂	1	0.0001	0.0050	0.9437
	temp × CO ₂	1	0.0005	0.0267	0.8707
	error	67	0.0212		
escape speed	temp	1	0.0034	0.0877	0.7680
	CO ₂	1	0.1897	4.8022	0.0318
	temp × CO ₂	1	0.3769	9.5418	0.0029
	error	67	2.6862		

There was a significant interaction between the high temperature and elevated CO₂ treatment for the mean prey escape speed (table 2; $F_{1,67} = 9.54$, $p = 0.002$) (figure 1*h*). Prey from the control temperature–control CO₂ treatment escaped significantly faster than those from the combined elevated CO₂ and high-temperature treatments (0.4 versus 0.26 m s⁻¹).

4. Discussion

The oceans are warming and seawater $p\text{CO}_2$ is rising due to increasing anthropogenic CO₂ emissions, with these environmental changes potentially having antagonistic, additive or synergistic effects on the physiology and behaviour of marine animals [4,6,9]. In this study, we show that short-term exposure to elevated CO₂ and high temperature led to pronounced changes in predator–prey interactions of reef fishes. In terms of the prey, high temperature had the strongest effect on reactivity (i.e. RD and ALT) and locomotory performance. However, the interaction between CO₂ and temperature was complex, resulting in a mildly additive response with CO₂ exposure slightly increasing the effect of temperature for some performance traits.

Prey exposed to high temperatures displayed a decrease in reactivity with an attenuation of RD and an increase in

ALT. The fast kinematics of escape responses are likely to be under the control of Mauthner cells, which are triggered as a reaction to the fast approach of a predator [37], with reactivity a crucial factor for predator evasion [38]. Our findings are in line with those of Webb & Zhang [19], who found that the RD and ALT of prey (goldfish, *Carassius auratus*) to an attacking predator (rainbow trout, *Oncorhynchus mykiss*) deteriorated following acute exposure to high temperatures, contributing to increased predator success. Exposure to high temperatures can lead to changes in the physiology of the Mauthner cell circuit, resulting in differences in the balance between the excitatory and inhibitory transmission onto the Mauthner cell [17].

Our results also showed that directionality was impaired in prey fish exposed to high temperatures. Directionality is an important trait, with an early mistake—such as turning towards a predator—likely to be costly [38]. Furthermore, the score of directionality is a good indicator of whether there is neural impairment associated with a treatment. The direction of the fast-start indicates whether the right or left Mauthner cell has been fired [37]. Typically, if the stimulus is on the right, the Mauthner cell that fires is also on the right, meaning that the muscle that contracts will be the contralateral one (i.e. on the left), leading to an ‘away’ response. In this study, prey exposed to higher temperatures tended to

show lower directional discrimination of the predator by exhibiting directionality that was not different from random (i.e. 50% away and 50% towards the stimulus). By contrast, individuals that were exposed to control conditions exhibited directionality that was different from random and significantly away from the threat. We also observed both an independent effect of elevated CO₂ and an effect due to the combined exposure (CO₂ and temperature) on directionality, with both groups turning away from or towards the predator randomly. Directionality was one of the few escape variables impaired by CO₂ exposure. The increase in capture success from 5% in the control treatment to 22% in the elevated CO₂ treatment, suggests that directionality is a fundamental factor facilitating successful escapes.

Interestingly, we saw little influence of elevated CO₂ on prey reactivity with both RD and ALT at similar levels as in fish under control CO₂. This is a surprising result as earlier studies have demonstrated numerous behavioural effects of elevated CO₂ on coral reef fishes, including *P. wardi* [10,51,52]. It has been suggested that behavioural abnormalities associated with CO₂ exposure involve a malfunction of the GABA_A neurotransmitter, an impairment that is restored following treatment with the GABA_A antagonist gabazine [52]. GABA_A receptors are found throughout the Mauthner cell [53] and previous work with a closely related species (*Pomacentrus amboinensis*) hypothesizes that impairment of these receptors may be the cause of decreased prey reactivity and escape performance during a predator–prey interaction [32]. Alternatively, these conflicting responses may be due to species-specific differences; despite the close phylogenetic history with *P. wardi*, *P. amboinensis* is known to be particularly sensitive to increasing CO₂ [54,55]. Previous studies have found little effect of high CO₂ exposure on the physiological performance of most juvenile reef fishes [51,56]. The physiological tolerance of these species to high CO₂ may at least partly explain why there were limited effects of high CO₂ on escape kinematics in this study. Furthermore, recent work has shown that a change in behaviour in response to elevated CO₂ is not ubiquitous, with some temperate species showing little influence of elevated CO₂ exposure on behaviour [57].

We observed significant combined effects of temperature and CO₂ on prey escape speed. These changes may be due to reduced muscle power resulting from changes in the contractile properties of the swimming muscles [16] associated with increased temperature exposure, as well as changes in aerobic scope. Although not directly measured in this study, declining aerobic scope at higher temperatures may explain the reduction in locomotory performance. Despite fast starts being powered anaerobically, an energy debt has to be paid off by post-exercise oxygen consumption, requiring more energy than used initially, creating an energy deficit [58]. Fishes with reduced aerobic scope in high-temperature conditions may therefore show low performance escapes as an energy-saving strategy [13]. Similarly, previous work on fish exposed to hypoxia has shown that their escape (i.e. anaerobic) swimming performance is impaired [50].

The results for the predators are more complex to interpret, as capture success is determined by predator performance and motivation as well as the interplay between the predator and the prey. Predators exposed to high temperatures had a significant increase in capture success compared with the other three groups, which may have been due to two non-mutually

exclusive possibilities. First, prey responsiveness may have decreased, thereby making them more vulnerable. Second, there may have been an increase in the effectiveness of the predators, leading to an increase in attack and predation rates. Increasingly, high temperatures are known to increase metabolic rate [59], which in turn increases energetic demand. This may manifest as an increase in predator motivation to capture prey. As food availability was kept constant between the four treatment groups, this seems likely to be the cause of the increase in attack motivation. By contrast, Ferrari *et al.* [23] found that predation rates increased after the combined exposure to elevated CO₂ (995 μ atm) and high temperatures (31°C), yet failed to find an increase in predation rates in response to high-temperature exposure alone [23]. Despite the increase in capture success, we saw little influence of temperature on the locomotory performance of the predator, suggesting that *P. fuscus* may be unaffected by temperature increases such as those used here, potentially as a result of its recent thermal experience or evolutionary history. Alternatively, there may have been differential acclimation occurring between the predator and the prey, despite the consistent time spent in treatment between the two species. In contrast with earlier work by Allan *et al.* [32], there was little influence of elevated CO₂ on the locomotory performance on the predator despite observing a fourfold increase in capture success compared with controls. This is consistent with the physiological tolerance to high CO₂ in *P. fuscus* reported by Couturier *et al.* [56].

We observed a strong effect of short-term temperature exposure on predator–prey interactions. This may lead to changes in the replenishment of recruits to coral reefs. The temperature used in our experiment was 3°C higher than the control October temperature of 27°C; however, both species (*Po. wardi* and *Ps. fuscus*) do experience temperatures around 30°C in the middle of summer. In Australia, October is the start of the breeding season and settling recruits experience a severe predation-induced bottleneck at this vulnerable time. Therefore, any changes in the numbers of fish that successfully settle could lead to changes in community structure. Examining interactions between species in response to environmental stressors can be complex due to behavioural subtleties that may not be captured. Only focusing on the responses of individual species is likely to provide an insufficient basis from which to gain an understanding of long-term responses to climate change in complex ecological environments, such as coral reefs [60] due to differing tolerances to environmental perturbations [20,22].

While this study examined short-term exposure to elevated temperature, the timeframe is relevant to exposure of new recruits on the reef. Furthermore, there is little evidence to suggest that reef fishes acclimate to elevated temperatures over longer durations (7–22 days) [61]. However, there is evidence for transgenerational thermal acclimation in reef fishes [62] and evidence for heritable variation in the metabolic responses of juvenile reef fishes to higher temperatures that could aid them in adapting to future warming [63]. Consequently, future studies will also need to consider the potential for adaptation. Our results, combining escape performance, attack behaviour and mortality rates, suggest that increasing CO₂ and warming may lead to complex changes in the predator–prey relationships of coral reef communities. The novelty is that this study examines a crucially important process that is fundamental to all food-webs: predation by a predator, and avoidance by a prey. To date, most studies have only examined physiological or performance aspects of

isolated species and have not explored the interactions among species. Understanding how organisms and ecosystems respond to key environmental drivers concurrently remains a priority for science, management and conservation, and this study provides the foundation for studies of other predator–prey interactions.

Ethics. Methods were approved by the Great Barrier Reef Marine Park Authority. All experimental protocols were approved under the James Cook University ethics guidelines (Approval: A2080).

Data accessibility. This article has no additional data.

References

- Collins M *et al.* 2013 *Climate Change: The Physical Science Basis. Contribution of Working Group I to the Fifth Assessment Report of the Intergovernmental Panel on Climate Change*. Cambridge, UK: Cambridge University Press.
- Doney SC, Fabry VJ, Feely RA, Kleypas JA. 2009 Ocean acidification: the other CO₂ problem. *Annu. Rev. Mar. Sci.* **1**, 169–192. (doi:10.1146/annurev.marine.010908.163834)
- Lüthi D *et al.* 2008 High-resolution carbon dioxide concentration record 650,000–800,000 years before present. *Nature* **453**, 379–382. (doi:10.1038/nature06949)
- Kroeker KJ, Kordas RL, Crim R, Hendriks IE, Ramajo L, Singh GS, Duarte CM, Gattuso J-P. 2013 Impacts of ocean acidification on marine organisms: quantifying sensitivities and interaction with warming. *Glob. Change Biol.* **19**, 1884–1896. (doi:10.1111/gcb.12179)
- Gaylord B *et al.* 2015 Ocean acidification through the lens of ecological theory. *Ecology* **96**, 3–15. (doi:10.1890/14-0802.1)
- Heuer RM, Grosell M. 2014 Physiological impacts of elevated carbon dioxide and ocean acidification on fish. *Am. J. Physiol. Integr. Comp. Physiol.* **307**, R1061–R1084. (doi:10.1152/ajpregu.00064.2014)
- Riebesell U, Gattuso J-P. 2015 Lessons learned from ocean acidification research. *Nat. Clim. Change* **5**, 12–14. (doi:10.1038/ndimate2456)
- Briffa M, de la Haye K, Munday PL. 2012 High CO₂ and marine animal behaviour—potential mechanisms and ecological consequences. *Mar. Pollut. Bull.* **64**, 1519–1528. (doi:10.1016/j.marpolbul.2012.05.032)
- Nagelkerken I, Munday PL. 2016 Animal behaviour shapes the ecological effects of ocean acidification and warming: moving from individual to community-level responses. *Glob. Change Biol.* **22**, 974–989. (doi:10.1111/gcb.13167)
- Munday PL, Dixon DL, McCormick MI, Meekan M, Ferrari MCO, Chivers DP. 2010 Replenishment of fish populations is threatened by ocean acidification. *Proc. Natl Acad. Sci. USA* **107**, 12 930–12 934. (doi:10.1073/pnas.1004519107)
- Chivers DP, McCormick MI, Nilsson GE, Munday PL, Watson S-A, Meekan MG, Mitchell MD, Corkill KC, Ferrari MCO. 2014 Impaired learning of predators and lower prey survival under elevated CO₂: a consequence of neurotransmitter interference. *Glob. Change Biol.* **20**, 515–522. (doi:10.1111/gcb.12291)
- Farrell AP. 2002 Cardiorespiratory performance in salmonids during exercise at high temperature: insights into cardiovascular design limitations in fishes. *Comp. Biochem. Physiol. A Mol. Integr. Physiol.* **132**, 797–810. (doi:10.1016/S1095-6433(02)00049-1)
- Johansen JL, Jones GP. 2011 Increasing ocean temperature reduces the metabolic performance and swimming ability of coral reef damselfishes. *Glob. Change Biol.* **17**, 2971–2979. (doi:10.1111/j.1365-2486.2011.02436.x)
- Eliason EJ *et al.* 2011 Differences in thermal tolerance among Sockeye salmon populations. *Science* **332**, 109–112. (doi:10.1126/science.1199158)
- Hanel R, Wieser W. 1196 Growth of swimming muscles and its metabolic cost in larvae of whitefish at different temperatures. *J. Fish Biol.* **48**, 937–951. (doi:10.1111/j.1095-8649.1996.tb01488.x)
- Wakeling JM. 2006 Fast-start mechanics. In *Fish biomechanics: fish physiology* (eds RE Shadwick, GV Lauder), pp. 333–368. San Diego, CA: Academic Press.
- Szabo TM, Brookings T, Preuss T, Faber DS. 2008 Effects of temperature acclimation on a central neural circuit and its behavioral output. *J. Neurophysiol.* **100**, 2997–3008. (doi:10.1152/jn.91033.2008)
- Biro PA, Beckmann C, Stamps JA. 2010 Small within-day increases in temperature affects boldness and alters personality in coral reef fish. *Proc. R. Soc. B* **277**, 71–77. (doi:10.1098/rspb.2009.1346)
- Webb PW, Zhang H. 1994 The relationship between responsiveness and elusiveness of heat-shocked goldfish (*Carassius auratus*) to attacks by rainbow trout (*Oncorhynchus mykiss*). *Can. J. Zool.* **72**, 423–426. (doi:10.1139/z94-059)
- Allan BJM, Domenici P, Munday PL, McCormick MI. 2015 Feeling the heat: the effect of acute temperature changes on predator–prey interactions in coral reef fish. *Conserv. Physiol.* **3**, cov011. (doi:10.1093/conphys/cov011)
- Freitas V, Campos J, Fonds M, Van der Veer HW. 2007 Potential impact of temperature change on epibenthic predator–bivalve prey interactions in temperate estuaries. *J. Therm. Biol.* **32**, 328–340. (doi:10.1016/j.jtherbio.2007.04.004)
- Grigaltchik VS, Ward AJW, Seebacher F. 2012 Thermal acclimation of interactions: differential responses to temperature change alter predator–prey relationship. *Proc. R. Soc. B* **279**, 4058–4064. (doi:10.1098/rspb.2012.1277)
- Ferrari MCO, Munday PL, Rummer JL, McCormick MI, Corkill K, Watson S-A, Allan BJ, Meekan MG, Chivers DP. 2015 Interactive effects of ocean acidification and rising sea temperatures alter predation rate and predator selectivity in reef fish communities. *Glob. Change Biol.* **21**, 1848–1855. (doi:10.1111/gcb.12818)
- Domenici P, Allan BJM, Watson, S-A, McCormick MI, Munday PL. 2014 Shifting from right to left: the combined effect of elevated CO₂ and temperature on behavioural lateralization in a coral reef fish. *PLoS ONE* **9**, e87969. (doi:10.1371/journal.pone.0087969)
- Nowicki JP, Miller GM, Munday PL. 2012 Interactive effects of elevated temperature and CO₂ on foraging behavior of juvenile coral reef fish. *J. Exp. Mar. Biol. Ecol.* **412**, 46–51. (doi:10.1016/j.jembe.2011.10.020)
- Faleiro F *et al.* 2015 Seahorses under a changing ocean: the impact of warming and acidification on the behaviour and physiology of a poor-swimming bony-armoured fish. *Conserv. Physiol.* **3**, cov009. (doi:10.1093/conphys/cov009)
- Miller GM, Kroon FJ, Metcalfe S, Munday PL. 2014 Temperature is the evil twin: effects of increased temperature and ocean acidification on reproduction in a reef fish. *Ecol. Appl.* **25**, 603–620. (doi:10.1890/14-0559.1)
- Munday P, Crawley N, Nilsson G. 2009 Interacting effects of elevated temperature and ocean acidification on the aerobic performance of coral reef fishes. *Mar. Ecol. Prog. Ser.* **388**, 235–242. (doi:10.3354/meps08137)
- Crain CM, Kroeker K, Halpern BS. 2008 Interactive and cumulative effects of multiple human stressors in marine systems. *Ecol. Lett.* **11**, 1304–1315. (doi:10.1111/j.1461-0248.2008.01253.x)
- Kneitel JM, Chase JM. 2004 Trade-offs in community ecology: linking spatial scales and species coexistence. *Ecol. Lett.* **7**, 69–80. (doi:10.1046/j.1461-0248.2003.00551.x)

31. Almany GR, Webster MS. 2006 The predation gauntlet: early post-settlement mortality in reef fishes. *Coral Reefs* **25**, 19–22. (doi:10.1007/s00338-005-0044-y)
32. Allan BJM, Domenici P, McCormick MI, Watson S-A, Munday PL. 2013 Elevated CO₂ affects predator–prey interactions through altered performance. *PLoS ONE* **8**, e58520. (doi:10.1371/journal.pone.0058520)
33. Green BS, McCormick MI. 2005 Maternal and paternal effects determine size, growth and performance in larvae of a tropical reef fish. *Mar. Ecol. Prog. Ser.* **289**, 263–272. (doi:10.3354/meps289263)
34. Holmes T, McCormick M. 2010 Size-selectivity of predatory reef fish on juvenile prey. *Mar. Ecol. Prog. Ser.* **399**, 273–283. (doi:10.3354/meps08337)
35. Domenici P, Blake R. 1997 The kinematics and performance of fish fast-start swimming. *J. Exp. Biol.* **200**, 1165–1178.
36. Rome LC, Funke RP, Alexander RM, Lutz G, Aldridge H, Scott F, Freadman M. 1998 Why animals have different muscle fibre types. *Nature* **335**, 824–827. (doi:10.1038/335824a0)
37. Eaton RC, Lee RKK, Foreman MB. 2001 The Mauthner cell and other identified neurons of the brainstem escape network of fish. *Prog. Neurobiol.* **63**, 467–485. (doi:10.1016/S0301-0082(00)00047-2)
38. Domenici P. 2010 Context-dependent variability in the components of fish escape response: integrating locomotor performance and behavior. *J. Exp. Zool. Part. Ecol. Genet. Physiol.* **313A**, 59–79. (doi:10.1002/jez.580)
39. Cripps IL, Munday PL, McCormick MI. 2011 Ocean acidification affects prey detection by a predatory reef fish. *PLoS ONE* **6**, e22736. (doi:10.1371/journal.pone.0022736)
40. Feeney WE, Lönnstedt OM, Bosiger Y, Martin J, Jones GP, Rowe RJ, McCormick MI. 2012 High rate of prey consumption in a small predatory fish on coral reefs. *Coral Reefs* **31**, 909–918. (doi:10.1007/s00338-012-0894-z)
41. Munday PL, Watson SA, Chung WS, Marshall NJ, Nilsson GE. 2014 Response to ‘The importance of accurate CO₂ dosing and measurement in ocean acidification studies’. *J. Exp. Biol.* **217**, 1828–1829. (doi:10.1242/jeb.105890)
42. Hari P, Pumpanen J, Huotari J, Kolari P, Grace J, Vesala T, Ojala A. 2008 High-frequency measurements of productivity of planktonic algae using rugged nondispersive infrared carbon dioxide probes. *Limnol. Ocean. Methods* **6**, 347–354. (doi:10.4319/lom.2008.6.347)
43. Pierrot D, Lewis E, Wallace D. 2006 *MS excel Program Developed for CO₂ System Calculations. ORNL/CDIAC-105a*. Oak Ridge, TN: Carbon Dioxide Information Analysis Center, Oak Ridge National Laboratory US Department of Energy.
44. Mehrbach C, Culbertson C, Hawley J, Pytkowicz R. 1973 Measurement of the apparent dissociation constants of carbonic acid in seawater at atmospheric pressure. *Limnol. Oceanogr.* **1973**, 897–907. (doi:10.4319/lo.1973.18.6.0897)
45. Dickson AG, Millero FJ. 1987 A comparison of the equilibrium constants for the dissociation of carbonic acid in seawater media. *Deep-Sea Res.* **34**, 1733–1743. (doi:10.1016/0198-0149(87)90021-5)
46. Webb PW. 1976 The effect of size on the fast-start performance of rainbow trout *Salmo gairdneri*, and a consideration of piscivorous predator–prey interactions. *J. Exp. Biol.* **65**, 157–177.
47. Webb PW. 1978 Fast-start performance and body form in seven species of teleost fish. *J. Exp. Biol.* **74**, 211–226.
48. Munday PL, Welch M, Allan BJM, Watson S-A, McMahon S, McCormick MI. 2016 Effects of elevated CO₂ on predator avoidance behaviour by reef fishes is not altered by experimental test water. *PeerJ* **4**, e2501. (doi:10.7717/peerj.2501)
49. Dill LM. 1974 The escape response of the zebra danio (*Brachydanio rerio*) II. The effect of experience. *Anim. Behav.* **22**, 723–730. (doi:10.1016/S0003-3472(74)80023-0)
50. Domenici P, Lefrancois C, Shingles A. 2007 Hypoxia and the antipredator behaviours of fishes. *Phil. Trans. R. Soc. B* **362**, 2105–2121. (doi:10.1098/rstb.2007.2103)
51. Munday PL, Cheal AJ, Dixon DL, Rummer JL, Fabricius KE. 2014 Behavioural impairment in reef fishes caused by ocean acidification at CO₂ seeps. *Nat. Clim. Change* **4**, 487–492. (doi:10.1038/nclimate2195)
52. Nilsson GE, Dixon DL, Domenici P, McCormick MI, Sorensen C, Watson S-A, Munday PL. 2012 Near-future carbon dioxide levels alter fish behaviour by interfering with neurotransmitter function. *Nat. Clim. Change* **2**, 201–204. (doi:10.1038/nclimate1352)
53. Diamond J, Huxley AF. 1968 The activation and distribution of GABA and L-glutamate receptors on Goldfish Mauthner neurons: an analysis of dendritic remote inhibition. *J. Physiol.* **194**, 669–723. (doi:10.1113/jphysiol.1968.sp008432)
54. Ferrari MCO *et al.* 2011 Intrageneric variation in antipredator responses of coral reef fishes affected by ocean acidification: implications for climate change projections on marine communities. *Glob. Change Biol.* **17**, 2980–2986. (doi:10.1111/j.1365-2486.2011.02439.x)
55. McCormick MI, Watson S-A, Munday PL. 2013 Ocean acidification reverses competition for space as habitats degrade. *Sci. Rep.* **3**, 3280. (doi:10.1038/srep03280)
56. Couturier CS, Stecyk JAW, Rummer JL, Munday PL, Nilsson GE. 2013 Species-specific effects of near-future CO₂ on the respiratory performance of two tropical prey fish and their predator. *Comp. Biochem. Physiol. A Mol. Integr. Physiol.* **166**, 482–489. (doi:10.1016/j.cbpa.2013.07.025)
57. Jutfelt F, Hedgarde M. 2013 Atlantic cod actively avoid CO₂ and predator odour, even after long-term CO₂ exposure. *Front. Zool.* **10**, 81. (doi:10.1186/1742-9994-10-81)
58. Moyes CD, Schulte PM, Andwest TG. 1993 Burst exercise recovery metabolism in fish white muscle. In *Surviving hypoxia: mechanisms of control and adaptation* (ed. PW Hochachka), pp. 527–539. Boca Raton, FL: CRC Press, Inc.
59. Pörtner HO, Farrell AP. 2008 Physiology and climate change. *Science* **322**, 690–692. (doi:10.1126/science.1163156)
60. Harley CGD. 2011 Climate change, keystone predation, and biodiversity loss. *Science* **334**, 1124. (doi:10.1126/science.1210199)
61. Nilsson GE, Östlund-Nilsson S, Munday PL. 2010 Effects of elevated temperature on coral reef fishes: loss of hypoxia tolerance and inability to acclimate. *Comp. Biochem. Physiol. A Mol. Integr. Physiol.* **156**, 389–393. (doi:10.1016/j.cbpa.2010.03.009)
62. Donelson JM, Munday PL, McCormick MI, Pitcher CR. 2011 Rapid transgenerational acclimation of a tropical reef fish to climate change. *Nat. Clim. Change* **2**, 30–32. (doi:10.1038/nclimate1323)
63. Munday PL, Donelson JM, Domingos JM. 2017 Potential for adaptation to climate change in a coral reef fish. *Glob. Change Biol.* **23**, 307–317. (doi:10.1111/gcb.13419)